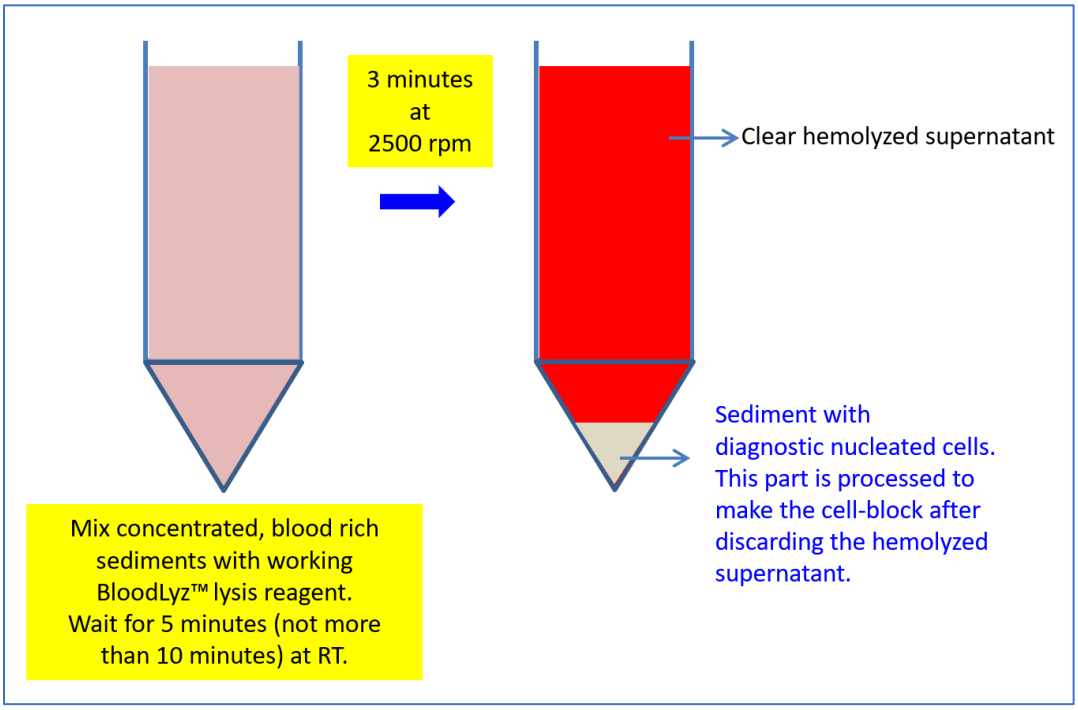
Summary protocol for BloodLyz™

1. For preparing working BloodLyz™ reagent, dilute 5 mL of concentrated 10X BloodLyz™ reagent provided in the tube by adding 45 mL of distilled water at room temperature up to the 50 mL mark.
2. Concentrate the specimen by centrifuging at 1250 RCF (2500 rpm on a centrifuge with rotor diameter of 11 cm) for 5 minutes. Decant supernatant by carefully removing the supernatant with a pipette.
3. Resuspend the sediments in the scant residual supernatant remaining after decanting.
4. Add working (diluted) BloodLyz™ to the blood contaminated concentrated sediment and disperse all the sediments in it (use up to 5 mL sediments for all 50 mL working BloodLyz™. If the sediments are less in quantity, you may economize and use a ratio of 0.5 mL sediment to 5 mL *working* (diluted) BloodLyz™. Extra unused working BloodLyz™ reagent may be stored at 2-8^{o}C and used within 7 days).
5. Cap securely and mix well by inverting gently a few times.
6. Wait for 5 minutes (not more than 10 minutes) at room temperature.
7. Centrifuge immediately at 2500 rpm for 3 minutes at room temperature.
8. Decant the reddish transparent hemolyzed supernatant.
9. Resuspend the nucleated cells in the whitish sediments and procced with cell-blocking protocol (without significant delay) using the Nano NextGen CelBloking™ (NGCB) Kit by adding the RBC-depleted concentrated sediment to the Nano NGCB unit.



Schematic summarizing the procedure for processing of blood contaminated cytology specimens with BloodLyz™ to nullify the problems related to red blood cell contamination.